

# Soybean Host Control of Nodulation by Strains of *Bradyrhizobium*

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Perry B. Cregan, USDA, ARS, Beltsville, MD 20705

Patrick E. Elia, USDA, ARS, Beltsville, MD 20705

Qijian Song, USDA, ARS, Beltsville, MD 20705

Fawzy Hashem, Univ. of MD Eastern Shore, Princess Anne, MD 21853

Randall L. Nelson, USDA, ARS, Urbana, IL 61801

Thomas E. Carter, USDA, ARS, Raleigh, NC 27607

Peter van Berkum, USDA, ARS, Beltsville, MD 20705

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# Background Information

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- ◆ Soybean roots nodulate and form a nitrogen fixing symbiosis with “rhizobia” that are classified as *Bradyrhizobium japonicum* and *B. elkanii*. Bacterial induced chlorosis by *B. elkanii*
  - ◆ Variability in the effectiveness of N<sub>2</sub> fixation among *B. japonicum* strains and the competitiveness of *B. japonicum* in U.S. soils
  - ◆ *B. japonicum* of serogroup USDA 123 occupy a large proportion of nodules of soybeans grown in northern Midwestern soils.
  - ◆ N<sub>2</sub> fixation and seed yield of serogroup USDA 123 strains versus other *B. japonicum*
  - ◆ Research to use the soybean host to reduce nodulation by indigenous serocluster 123 *B. japonicum* strains
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# Background Information

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**Soybean roots nodulate and form a nitrogen fixing symbiosis with bacteria that are classified as *Bradyrhizobium japonicum* and *B. elkanii***

- ◆ **Jordan (1982): Renaming of the slow-growing Legume nodulating bacteria based upon growth rate in yeast extract-mannitol medium**

The soybean nodulating bacteria previously referred to as “*Rhizobium japonicum*” were renamed “*Bradyrhizobium japonicum*”

- ◆ **Kuykendall et al. (1992): Based upon DNA homology, RFLP analysis and other traits divided the soybean *Bradyrhizobia* into two groups**

*Bradyrhizobium elkanii* was the name given to a portion of the soybean-nodulating Bradyrhizobia stains. These are the stains that had previously been shown to produce *Rhizobium*-induced chlorosis on susceptible soybean genotypes.

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# Background Information

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## Bacterial-induced chlorosis by strains of *B.*

- ◆ Erdman et al. (1997): *Rhizobium*-induced chlorosis produced by strains of *B. elkanii* seen in Southern U.S. soybean fields especially in the cultivar Lee  
“Seed yields are not measurably reduced” by *Rhizobium*-induced chlorosis
  - ◆ Johnson and Means (1960): *Rhizobium*-induced chlorosis varied in soybean genotype x *Rhizobium* strain combinations grown in GH and growth chamber trials  
“When present, severe *Rhizobium*-induced chlorosis, the relationship between bacteria and host can hardly be regarded as symbiotic”
  - ◆ Fuhrmann (1990): 18% of *Bradyrhizobium* isolates from Delaware soils produced Rhizobitoxin chlorosis symptoms  
In a greenhouse pot experiment plant N content significantly lower with Rhizobitoxin-producing strains
  - ◆ Teaney and Fuhrmann (1992): In the absence of discernable foliar chlorosis, the effect of nodulation by Rhizobitoxin-producing *Bradyrhizobium* strains in minimal
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# Background Information

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## Variability in the effectiveness of N<sub>2</sub> fixation has been reported among strains of *B.*

- ◆ **Abel *japonicum* (1964)**: The seed yield of “Lee” soybean grown on *B. japonicum*-free soil with 23 *B. japonicum* inoculation treatments and an uninoculated control treatments ranged from 1564.9 kg/ha to 2968.4 kg/ha (USDA Strain 110). Uninoculated control yielded 1477.5 kg/ha
  - ◆ **Caldwell and Vest (1970)**: Yield trails were conducted for three years on *B. japonicum* free soil using 28 *B. japonicum* strains and two commercial soybean treatments. No significant differences were found between strains. No significant strain x cultivar interactions were detected.
  - ◆ **Ham (1980) and Chamber *et al.* (1983)**: Yield tests conducted on *B. japonicum*-free soil with inoculation with various *B. japonicum* strains. Superior quality strains such as USDA 110 and USDA 138 produced superior seed yields.
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# Background Information

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## The competitiveness of soybean nodulating *Bradyrhizobium* in U.S. soils and nodulation by inoculant strains

- ◆ Johnson et al. (1965): Inoculated Maryland and Iowa field plots with 25, 100, 200, 400 and 800 times the normal inoculation rate + other additional treatments. Inoculation did sometimes increase the level of inoculant strains in nodules but standard inoculation rates did not
- ◆ Weaver and Frederick (1974): Inoculated Iowa field plots with 1.0 ml per cm of row using liquid cultures with up to  $3 \times 10^8$  cells/ml of *B. japonicum* USDA 138. “Inoculum densities are to form 50% or more of nodules an inoculum rate of at least 1,000 times the soil population (per g soil) must be used”
- ◆ Kvien et al. (1981): Examined the competitiveness of an applied inoculum strain against the indigenous *Bradyrhizobium* at Minnesota field sites in which serotype strains USDA 123 predominated  
Highly significant soybean genotype differences in the recovery of inoculant strain USDA 110 as well as large differences in recovery between field sites and years



# Background Information

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***B. japonicum* of serogroup USDA 123 occupy a large proportion of nodules of soybeans grown in northern Midwestern soils**

- ◆ Damirgi et al. (1987): Determined the serotype of *B. japonicum* in the nodules of soybeans collected from various soil types “in widely separated areas of Iowa”  
Nodules obtained were in one of four serogroups (USDA 123, 135, 31 and 3). Serogroup 123 was dominant averaging 52% across soil types.
  - ◆ Ellis et al. (1984): Used *B. japonicum* inoculation rates up to 50 times the normal rate and determined the % of nodules with inoculant strain vs. indigenous serogroup 123  
95.2% of nodules were occupied with the indigenous serogroup 123 strain.
  - ◆ Moawad et al. (1984): Measured rhizosphere densities of serogroup USDA 110, 123 and 138 and nodule occupancy by these serogroup strains on 3 soybean genotypes  
Although serogroup 123 gave no evidence of dominance in early host rhizospheres it clearly dominated in nodule composition, occupying 60 to 100% of the nodules.”
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# Background Information

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**Nodulation by indigenous *B. japonicum* of serogroup USDA 123 does not provide highly effective N<sub>2</sub> fixation or the highest seed yields**

- ◆ **Caldwell and Vest (1970):** 28 *B. japonicum* strains and 2 commercial inocula used in replicated yield trials on *B. japonicum*-free soil for 3 years  
The yield of cultivars inoculated with USDA 123 ranked 16th among the 30 strains.
- ◆ **Ham et al. (1980):** Seed yield of three soybean cultivars grown on *B. japonicum*-free soil with 7 *B. japonicum* strains  
Seed yield with USDA 123 ranked last among the 7 strains.
- ◆ **Kvien et al. (1981):** Yield trials with 12 genotypes conducted on typical Minnesota soils with and without inoculation. Nodule occupancy determined for all treatments.  
In the uninoculated treatments 95% of the nodules were occupied with serogroup 123 strains. “The 12 soybean lines responded with yield increases whenever 50% or more of the nodules were formed by the inoculant strains.”



# Background Information

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Could the genetics of the soybean host be manipulated to stop nodulation by indigenous serogroup 123 *B. japonicum*?

- ◆ Caldwell (1966): Reported the single dominant gene *Rj2* that restricts the nodulation of all available strains of the USDA 122 and c1 serogroups.
- ◆ Vest (1970): Reported the single dominant gene *Rj3* that restricts the nodulation of strain USDA 33 but no other strains that are serologically related.
- ◆ Vest and Caldwell (1972): Reported the single dominant gene *Rj4* that restricts nodulation with strain USDA 61.

Does a soybean genotype exist that would restrict nodulation by strains of indigenous serogroup USDA 123 while allowing normal nodulation and N<sub>2</sub> fixation with ~~other more effective strains?~~

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# Background Information

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## Identification of soybean genotypes that restrict the nodulation of strain USDA 123 and other serologically

- ◆ **related strains (1986): 1287 soybean cultivars and Plant Introductions were screened in the greenhouse for nodulation with strain USDA 123.**
    - ∞ Two genotypes (PI371607 and PI377578) were identified that restricted the nodulation of strain USDA 123.
    - ∞ In competition studies with USDA 123 vs. inoculant quality strains, greater than 75% of the nodules on the control genotype Williams were occupied with USDA 123 while less than 10% of the nodules of the PI genotypes contained USDA 123.
  - ◆ **Keyser and Cregan (1987): 20 field isolates of serogroup 123 strains tested for nodulation of the two USDA 123-restricting PI genotypes and Williams**
    - ∞ The nodulation of only 4 of the field isolates were restricted by the PI genotypes these were serogroup 123 strains.
    - ∞ The strains that were not restricted were determined to be USDA 127 and USDA 129 serotypes – members of “serocluster 123”(as defined by Schmidt et al. (1986).
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# Background Information

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## Identification of soybean genotypes that restrict the nodulation of previously unrestricted “serocluster” 123

### ◆ *B. japonicum* strains

◆ Cregan, Keyser & Sadowsky (1989): 850 soybean cultivars and Plant Introductions were screened in the greenhouse for nodulation with serocluster 123 strain MN1-1c (serotype 127).

- ∞ Two genotypes (PI417566 and PI283326) were identified that restricted nodulation of strain MN1-1c.
- ∞ The PI genotypes also restricted the nodulation of inoculant quality strains USDA 110 and USDA 138. PI283326 restricted the nodulation of a number of other strains.
- ∞ In competition studies with MN1-1c vs. inoculant quality strain CB1809, over 90% of the nodules on the control genotype, Williams, were occupied with MN1-1c while less than 10% with CB1809. 70% of the nodules on PI417566 were occupied by CB1809.



# Background Information

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## Observations and questions raised by previous research regarding soybean host controlled strain-specific nodulation restriction of *Bradyrhizobium* strains

- ◆ There are apparently a number or perhaps many different genes that control strain specific nodulation restriction: *Rj2*, *Rj3*, *Rj4*, strain USDA 123 specific nodulation restriction, strain MN1-c specific nodulation restriction
- ◆ Is there really any relationship between *B. japonicum* serology and nodulation restriction?
- ◆ What strains/serotypes actually form nodules when genotypes with various nodulation restricting characteristics are grown in soils in which soybeans are normally grown?



# Multilocus Sequence Typing (MLST) to Identify Genotypes of *Bradyrhizobium* and to Identify Nodule

Occupants  
van Berkum *et al.* 2012. MPMI 25:321-330

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- ◆ Based upon the sequence analysis of seven chromosomal loci in 190 soybean nodulating strains as well as Rhizobia and Bradyrhizobia of other legumes from the USDA National Rhizobium Resource Collection
  - ◆ The seven chromosomal loci: *asd*, *gapA*, *gyrB*, *ilvI*, *lepA*, *mdh* and *purC*
  - ◆ The sequence of an average of 425 bp from each of 7 chromosomal genes are used to define the allele present in each *Bradyrhizobium* strain
  - ◆ A combination of the alleles for each of the seven gene fragments is used to define the allelic profile or “GT” of each strain
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# Multi-location Field Trials to Analyze Soybean Host Control of Nodulation by Strains of *Bradyrhizobium*

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- ◆ Four locations: Illinois, North Carolina and two Maryland locations (Beltsville and Princess Anne)
  - ◆ Five soybean genotypes: Williams 82, Bragg, PI 371607, Peking and Evans in Randomized Complete Block experiments with 4 replications
  - ◆ The root systems of 4 plants from each rep removed from the soil at 60 days after emergence and washed to remove soil, stored frozen and sent to Beltsville.
  - ◆ Nodules removed from plants and bacteroids recovered from a random sample of 96 nodules per genotype-rep combination
  - ◆ Genomic DNA isolated from each bacteroid and MLST applied based upon the sequence analysis of gene fragments from *gapA*, *gyrB* and *mdh*
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# Nodule occupancy of *B. japonicum* vs. *B. elkanii*

Highly significant (< 0.0001) differences in the frequencies of *B. japonicum* and *B. elkanii* at the three locations:

	% Nodule Occupancy		
	<u>Beltsville</u>	<u>Illinois</u>	<u>N. Carolina</u>
<i>B. japonicum</i>	78.8 a	85.8 a	28.7 b
<i>B. elkanii</i>	21.2 b	14.2 b	71.3 a

No significant differences among genotypes in the frequencies of *B. japonicum* and *B. elkanii* at the three locations:

	<i>B. japonicum</i> % Nodule Occupancy			<i>B. elkanii</i> % Nodule Occupancy		
	<u>Beltsville</u>	<u>Illinois</u>	<u>N. Carolina</u>	<u>Beltsville</u>	<u>Illinois</u>	<u>N. Carolina</u>
Williams 82	84.9 a	89.5 a	31.1 a	15.1 a	10.5 a	68.9 a
Bragg	81.6 a	88.4 a	25.5 a	18.4 a	11.6 a	74.5 a
PI 371607	69.8 a	79.5 a	29.6 a	30.2 a	20.5 a	70.4 a



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# Nodule occupancy of *B. japonicum* strains at the Beltsville and Illinois locations

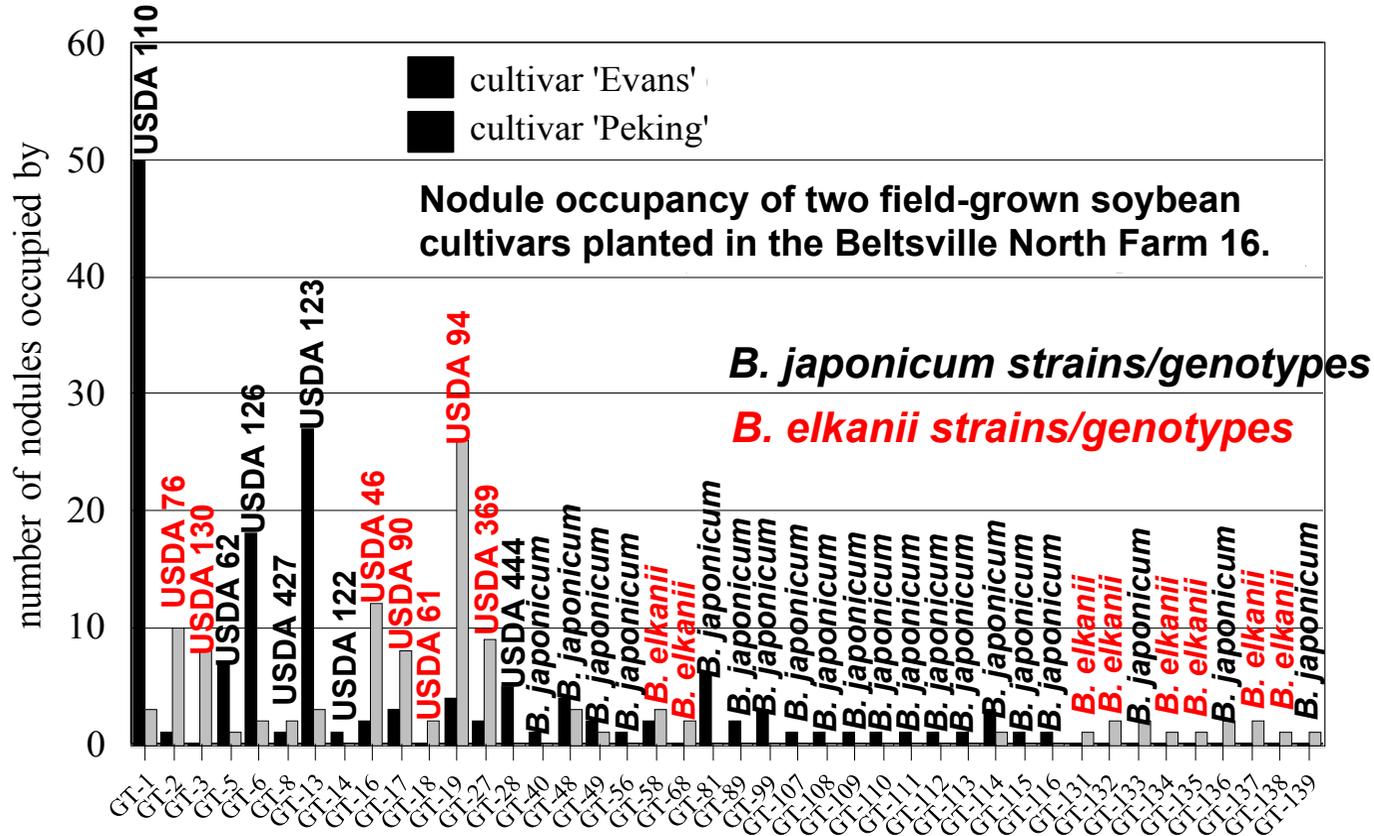
- ◆ There were no PI 371607 nodules occupied by *B. japonicum* USDA 123.

No significant differences among genotypes for nodule occupancy by inoculant quality strains USDA 110 or USDA 6

	Beltsville			Illinois		
	<u>% of <i>B. japonicum</i> nodules occupied by</u>			<u>% of <i>B. japonicum</i> nodules occupied by</u>		
	<u>USDA 123</u>	<u>USDA 110</u>	<u>USDA 6</u>	<u>USDA 123</u>	<u>USDA 110</u>	<u>USDA 6</u>
Williams 82	4.9 a	44.2 a	17.8 a	26.0 a	38.0 a	26.6 a
Bragg	2.6 a	72.9 a	20.6 a	33.3 a	33.3 a	36.3 a
PI 371607	0.0 a	74.7 a	6.3 a	0.0 b	53.8 a	12.0 a



# The impact of soybean genotype on nodule occupancy by *B. japonicum* vs. *B. elkanii*



	<u>% <i>B.j</i></u>	<u>%</u>
<i>B.e</i>		
Evans	92.2 a	
Peking	27.3 b	
		72.7 a



# Future research to understand and use soybean genetic control of nodulation to maximize the efficiency of N<sub>2</sub> fixation and enhance soybean yield

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- ◆ Multi-Locus Sequence Typing will allow a better understanding of the relationship between *B. japonicum* serology and restricted nodulation, assuming there is one.
- ◆ There is clearly much soybean genetic variation that impacts nodulation by a diversity of different *B. japonicum* genotypes/serotypes.
- ◆ The genetic control of genotype/serotype nodulation restriction can be determined.
- ◆ The molecular basis of host controlled nodulation restriction can be determined.
- ◆ With this knowledge can a novel plant-*Bradyrhizobium* recognition system be engineered that will only allow nodulation by a specific *Bradyrhizobium* with high symbiotic capacity?
- ◆ Will the reduction of nodulation by *B. elkanii* in favor of *B. japonicum* have a positive impact on N<sub>2</sub> fixation and seed yield?